



National Institute of Oceanography and Fisheries
Egyptian Journal of Aquatic Research

<http://ees.elsevier.com/ejar>
www.sciencedirect.com



FULL LENGTH ARTICLE

Seasonal fluctuations of phytoplankton community and physico-chemical parameters of the north western part of the Red Sea, Egypt



Mohamed Z. Nassar ^{a,*}, Hamdy R. Mohamed ^b, Hanan M. Khiray ^a,
 Sarah H. Rashedy ^a

^a National Institute of Oceanography and Fisheries, Egypt

^b Botany Department, Faculty of Science, South Valley University, Egypt

Received 7 September 2014; revised 3 November 2014; accepted 3 November 2014

Available online 2 December 2014

KEYWORDS

Phytoplankton;
 Species diversity;
 Physico-chemical
 parameters;
 Red Sea;
 Egypt

Abstract Phytoplankton community structure and some environmental parameters in the coastal water of the north western part of the Red Sea were studied seasonally during 2013. A total of 145 species were recorded with clear dominance of Bacillariophyceae, which formed about 76.4% of the total phytoplankton counts with annual average of 3654 cell/L and Dinophyceae (14.63%) with annual average of 700 cell/L. Other algal classes; like Cyanophyceae, Chlorophyceae, Euglenophyceae and Silicoflagellates sustained low counts, forming collectively about 9.0% of the total abundance of phytoplankton. Autumn was the most productive season recording an average of 5916 unit/L, followed by spring (average of 5282 unit/L) and winter (average of 4329 unit/L), while summer showed the lowest counts (average of 3607 unit/L). The species diversity fluctuated between 3.36 in the summer and 3.97 in autumn, with an annual average of 3.76.

The physico-chemical properties of surface water exhibited seasonal and spatial variations. The dissolved nitrate (0.07–2.27 μM), ammonium (1.82–8.80 μM), reactive silicate (0.41–5.22 μM) and water salinity (39.9–42.9‰) were the most effective factors that controlled the seasonal fluctuations of phytoplankton during 2013. The multiple regression model was: phytoplankton counts = 28,564 + 0.69 NO₃ + 0.284 NH₄ – 0.13 SiO₄ – 0.30 Salinity (M.R. = 0.91, *N* = 24 and *p* < 0.07). This equation could be applied in the future to predict the total phytoplankton counts in the coastal waters of the northern part of the Red Sea, Egypt.

© 2014 Hosting by Elsevier B.V. on behalf of National Institute of Oceanography and Fisheries. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/3.0/>).

Introduction

The Red Sea is a large marine ecosystem and it is an important economic and environmental asset (Longhurst, 2007 and Belkin, 2009). It is lying between the African and the Asian continental shelves, and is about 2250 km long (Fig. 1). At

* Corresponding author at: National Institute of Oceanography and Fisheries, P.O. Box 182, Suez, Egypt.

E-mail address: mnassar65@yahoo.com (M.Z. Nassar).

Peer review under responsibility of National Institute of Oceanography and Fisheries.

<http://dx.doi.org/10.1016/j.ejar.2014.11.002>

1687-4285 © 2014 Hosting by Elsevier B.V. on behalf of National Institute of Oceanography and Fisheries.

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/3.0/>).

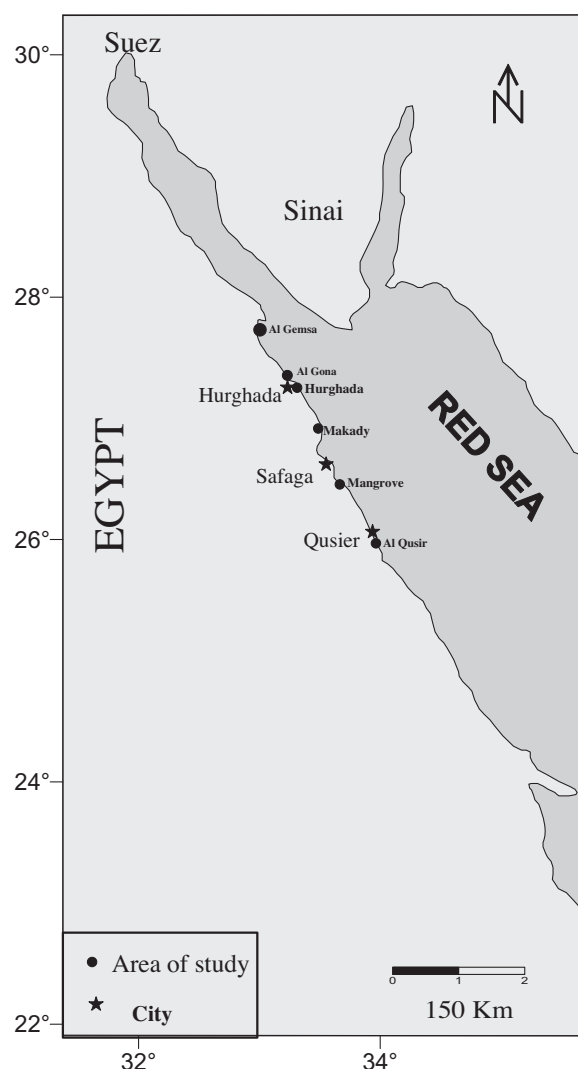


Figure 1 Positions of the sampling stations.

the northern extremity it is divided by the Sinai Peninsula into the Gulfs of Suez and Aqaba. The Suez Canal connects the Red Sea with the Mediterranean Sea, while the southern region exchanges waters with the open Indian Ocean through the Gulf of Aden and the Arabian Sea via the strait of Bab-el-Mandeb (Sofianos and Johns, 2002).

The Red Sea provides habitats for a wide range of marine species some of which are endemic (Baars et al., 1998) and can be considered a highly productive ecosystem. The pollution in the north western part of the Red Sea induced by the anthropogenic activities including oil spills and excessive loading of nutrients through addition of fertilizers and industrial wastewater and sewage has been reported (Abou-Aisha et al., 1995).

The phytoplankton plays an important role in the marine food web, biogeochemical cycle and climatic processes (Paerl et al., 2003 and Armbrust, 2004). In the oceanic waters of the central Red Sea, Halim (1969) reported 125 dinoflagellate species and 84 diatoms and Shaikh et al. (1986) detected 110 dinoflagellates and 137 diatoms. In the coastal waters of the Red Sea, El-Sherif and Abo El-Ezz (2000) examined the distribution of plankton at Taba, Sharm El-Sheikh, Hurghada and

Safaga on the Red Sea, recording 41 diatom species, 53 dinoflagellates, 10 cyanophytes and two chlorophytes. Sommer (2000) studied the relationship between larger nanophytoplankton and microphytoplankton and the nutrient limitation and grazer in the Gulf of Aqaba and the open northern Red Sea. AL-Qutob et al. (2002) followed the relationship between nitrite and phytoplankton in the Gulf of Aqaba. Deyab et al. (2004) recorded 200 phytoplankton species along the Suez Canal, Suez Gulf and the northern part of the Red Sea with the clear dominance of diatoms. Nassar (2007a) studied the phytoplankton dynamics in the coastal waters of Suez Gulf and recorded a total of 144 species of different groups, and Nassar (2007b) conducted a similar study on the phytoplankton abundance in the coastal waters of the Aqaba Gulf, recording 127 taxa. Also, Al-Najjar et al. (2007) studied the seasonal dynamics of phytoplankton in the Gulf of Aqaba. Toulbah et al. (2010) studied the phytoplankton community and physico-chemical characters of Jeddah coast, Red Sea. They reported that the coastal waters were found to be oligotrophic in some areas, while other areas were mesotrophic with high phytoplankton density. Madkour et al. (2010) studied the phytoplankton population along the southern part of Sinai Peninsula and the Gulfs of Suez and Aqaba. The phytoplankton population was fairly diversified (181 species) and comprised mainly two groups; dinoflagellates (116 species) and diatoms (60 species). There were relatively low variations in the phytoplankton composition in the study area. Spatial distribution of phytoplankton showed that Gulf of Suez differs in the dominant species and timing of abundance from both Gulf of Aqaba and the southern sites of Sinai Peninsula. Recently, Qurban et al. (2014) indicated that, the coastal waters in the Saudi Arabia of the northern Red Sea were oligotrophic and the primary production was strongly nitrogen-controlled.

Aim of work

The aim of the present study is to follow up the species composition and abundance of phytoplankton in the coastal water from Al Gemsha to Al Qusir along the north western part of the Red Sea in relation to the seasonal fluctuations of some physicochemical parameters.

Material and methods

The sample collection for phytoplankton study and physico-chemical measurements was carried out seasonally in winter (January), spring (April), summer (August) and autumn (November) during 2013 at six stations representing different ecological habitats along the northern Red Sea (Fig. 1). Al Gemsha (St.I) is located about 60 km north of Hurghada City and is subjected to low oil and sewage effluents of Al-Gemsha and the General Petroleum Companies, Al-Gona (St.II) lies at about 20 km north Hurghada City and near to the human and tourist activities, Hurghada (St.III) is found in front of the National Institute of Oceanography and Fisheries and is subjected to land filling and weak sewage effluents, Makady (St.IV) is located about 30 km south of Hurghada City and is relatively far from the pollution sources. Mangrove (St.V) is situated about 17 km south Safaga City (Pristine station) and Al Qusir (VI) is located about 140 km south of Hurghada City and is subjected to fishing activities and sewage impacts.

The pH value, water salinity and temperature were determined from the average of three readings at each station using the multiparameter instrument (Hanna Instruments, HI 9829). Dissolved oxygen was determined by Winkler's method (Strickland and Parsons, 1972) and the nutrient salts (NO_3 , NO_2 , NH_4 , PO_4 and SiO_4) were determined spectrophotometrically in μM according to the methods described by APHA (2005).

The species composition and standing crop of the phytoplankton were determined by the sedimentation method (Utermöhl, 1936) and expressed in cell or unit per liter, and the species identification was carried out following Peragallo and Peragallo (1908), El-Nayal (1935), Huber-Pestaluzzi (1938), Ghazzawi (1939), Cupp (1943), Prescott (1962), Bourrelly (1968), Ferguson Wood (1968), Stewart and Mattox (1975), Sourina (1986), Mizuno (1990) and Al-Kandari et al. (2009). However, the valid and accepted names of the phytoplankton species during the present study were according to the taxonomic database sites, like algaebase.com (ab), World Register of Marine Species (WoRMS), Canadian Register of Marine Species (CaRMS), Nordic Microalgae and Aquatic Protozoa (NOD) and Integrated Taxonomic Information System (ITIS).

The correlation matrices and the stepwise multiple regression analysis were applied using the program of STATISTICA Version 5. The species diversity (H') was calculated according to Shannon and Wiener (1963). The similarity matrix (S) and coefficient were calculated by the Bray and Curtis formula (Field et al., 1982) using PRIMER program Version 5.2.

Results and discussion

Physico-chemical parameters

Temperature

The surface water temperature is usually influenced by the intensity of solar radiation, evaporation and insulation and

the low temperature during monsoon. This could be due to strong sea breeze and cloudy sky (Behrenfeld et al., 2006). In this investigation, the costal water temperature varied between a minimum of 17.5 °C during winter at St.III and a maximum of 31.3 °C during summer at the same station and also at St.VI (Table 1). The autumn temperatures (21.1–24 °C) appeared to be suitable for the phytoplankton flourishing, where the phytoplankton standing crop reached its maximum abundance (average of 5916 unit/L). Similar observations were obtained by Nassar (2000) in the coastal water of the Suez Gulf. However, Boyce et al. (2010) indicated a decline in the marine phytoplankton biomass and primary productivity was found with increasing the sea surface temperatures. Also, the water warming usually increasing the reproduction rates and grazing activity of phytoplankton consumers (O'Connor et al., 2009).

pH

Marine phytoplankton in general is resistant to climate change in terms of ocean acidification and does not increase or decrease in its growth rate according to ecological relevant ranges of pH and CO_2 (Berge et al., 2010). The pH values were found on the alkaline side with a maximum of 8.80 at St.II during summer and a minimum of 7.80 at St.III during winter. The maximum phytoplankton standing crops during autumn was associated with the pH value of 8.05, supporting the observations of Toulibah et al. (2010) in the coastal water near Jeddah, Red Sea.

Water salinity

The salinity is the main physical parameter that can be attributed to the plankton diversity and acting as a limiting factor, which influences the distribution of plankton community (Sridhar et al., 2006). Our observations indicated salinity variations between 42.9‰ during summer at St.IV and 39.9‰ at St.III during autumn (Table 1). This high summer salinity may be attributed to the high evaporation rate as suggested

Table 1 Seasonal fluctuations of the physico-chemical parameters in the north western Red Sea during 2013.

Station	Season											
	Winter						Spring					
	I	II	III	IV	V	VI	I	II	III	IV	V	VI
Temp.	18.7	19.2	17.5	18.2	20.6	20.6	25.2	26.5	26.6	26.5	24.8	26.9
pH	8.18	8.00	7.80	8.00	7.90	7.90	7.90	7.87	7.90	7.90	7.90	7.90
Salinity	41.3	41.0	41.3	41.2	41.1	41.1	41.8	41.2	40.8	41.2	41.6	41.2
DO	6.2	7	7.1	7	6.9	6.7	7.1	6.7	7.0	6.5	6.9	7.8
PO_4	0.32	0.17	0.15	0.34	0.18	0.24	0.15	0.12	0.25	0.13	0.05	0.25
NO_3	0.45	0.42	1.47	0.66	0.39	1.21	1.03	0.54	1.78	0.40	0.75	1.52
NO_2	0.06	0.03	0.11	0.16	0.07	0.25	0.10	0.09	0.16	0.11	0.08	0.18
NH_4	4.69	3.69	3.95	3.44	2.23	5.22	3.22	2.82	3.35	3.52	2.32	4.06
SiO_4	0.97	1.77	3.42	2.65	2.06	1.39	1.6	1.63	1.91	1.55	1.60	1.46
Summer							Autumn					
Temp.	28.1	30.5	31.3	30.5	31.0	31.3	22.3	24.0	21.1	23.2	23.1	21.3
pH	8.60	8.80	8.65	8.6	8.73	8.60	8.00	7.90	7.90	8.30	8.10	8.12
Salinity	42.7	42.5	42.6	42.9	42.1	42.1	40.6	40.4	39.9	40.2	40.3	40.4
DO	6.4	6.7	7.8	7.5	6.2	6.3	6.4	6.7	8.2	7.9	6.7	8
PO_4	0.06	0.07	0.13	0.09	0.04	0.11	0.13	0.31	0.51	0.15	0.23	0.35
NO_3	0.36	0.79	1.05	0.16	0.07	0.73	0.44	1.01	2.27	1.19	0.68	1.35
NO_2	0.04	0.05	0.08	0.10	0.02	0.11	0.08	0.16	0.18	0.18	0.15	0.39
NH_4	8.80	5.05	4.25	3.96	3.06	6.21	2.69	2.21	3.18	2.22	1.82	2.58
SiO_4	1.92	2.21	3.35	3.88	5.22	2.69	0.62	1.49	0.41	1.02	1.03	0.83

Table 2 Annual average counts (unit/L) of the different phytoplankton classes and their frequency percentage during 2013.

Class	Station						Freq. %
	I	II	III	IV	V	VI	
Bacillariophyceae	3989	3321	5291	3262	2015	4047	76.4
Dinophyceae	690	700	1060	685	385	680	14.63
Cyanophyceae	145	105	315	120	220	275	4.12
Chlorophyceae	313	30	375	40	115	266	3.95
Silicoflagellates	40	55	30	55	0	15	0.69
Euglenophyceae	0	0	0	25	35	0	0.21
Total	5177	4211	7071	4187	2770	5283	100

Table 3 Seasonal variations of the different phytoplankton classes (unit/L) during 2013.

Class	Season				
	Winter	Spring	Summer	Autumn	Average
Bacillariophyceae	3371	4105	2587	4553	3654
Dinophyceae	626	653	596	925	700
Cyanophyceae	168	267	139	215	197
Chlorophyceae	117	217	239	186	189
Silicoflagellates	30	40	33	27	33
Euglenophyceae	17	0	13	10	10
Total	4329	5282	3607	5916	4783

Table 4 The mean counts of the dominant phytoplankton species (unit/L) in the north western Red Sea during 2013.

Class	Season				
	Winter	Spring	Summer	Autumn	Average
Bacillariophyceae (cell/L)					
<i>Climacospheia moniligera</i> Ehrenberg	130	123	460	137	213
<i>Cylindrotheca closterium</i> (Ehrenberg) Reimann & J.C. Lewin	130	277	150	217	194
<i>Chaetoceros simplex</i> Ostenfeld	250	248	99	149	187
<i>Chaetoceros curvisetus</i> Cleve	207	120	0	333	165
<i>Skeletonema costatum</i> (Greville) Cleve	117	140	240	137	159
<i>Chaetoceros lorenzianus</i> Grunow	73	167	123	273	159
<i>Nitzschia pungens</i> var. <i>atlantica</i> Cleve	107	150	97	123	119
<i>Licmophora flabellata</i> C. Agardh	103	97	103	100	101
<i>Leptocylindrus danicus</i> Cleve	93	117	60	80	88
<i>Asterionellopsis glacialis</i> (F. Castracane) F.E. Round	113	60	60	113	87
<i>Coscinodiscus granii</i> Gough	103	120	40	80	86
<i>Navicula cancellata</i> Donkin	113	77	67	53	78
<i>Proboscia alata</i> (Brightwell) Sündstrom	63	170	27	53	78
Dinophyceae (cell/L)					
<i>Ceratium furca</i> (Ehrenberg) Claparède & Lachmann	75	77	68	180	100
Chlorophyceae (unit/L)					
<i>Chlorella salina</i> Butcher	67	0	133	96	74
<i>Dictyosphaerium pulchellum</i> H.C.Wood (accepted name:	27	147	30	50	64
<i>Mucidosphaerium pulchellum</i> (H.C.Wood) C. Bock, Proschold & Krienitz					

by Khomayis (2002) and Schumann et al. (2006). The water salinity was one of the environmental factors affecting the phytoplankton abundance in the study area, where the highest abundance of phytoplankton during autumn coincided with low salinity values, that can be confirmed by the negative correlation between salinity and the phytoplankton counts ($r = 0.59$ at $p < 0.05$ & $N = 24$) as shown in (Table 5).

Dissolved oxygen (DO)

The dissolved oxygen in water is usually depending on its temperature and salinity. It is also depending on a considerable degree on the quantity of organic matter present in the aquatic environment. If the decomposition of organic matter is in great proportion, it will absorb too much of the dissolved oxygen in water (Shakweer, 2003). The results in Table 1, indicate that

Table 5 The correlations between the physico-chemical parameters and the total counts of phytoplankton in the north western Red Sea during 2013.

	Phyto.	Temp.	pH	Salinity	DO	NO ₃	NO ₂	NH ₄	PO ₄	SiO ₄
Phyto.	1.00									
Temp.	− 0.30	1.00								
pH	−0.42	0.70	1.00							
Salinity	−0.59	0.67	0.71	1.00						
DO	0.55	−0.16	−0.13	−0.32	1.00					
NO ₃	0.86	−0.23	−0.40	−0.52	0.65	1.00				
NO ₂	0.51	−0.30	−0.33	−0.54	0.53	0.60	1.00			
NH ₄	−0.00	0.31	0.48	0.57	−0.28	−0.13	−0.24	1.00		
PO ₄	0.60	0.58	0.47	−0.69	0.49	0.62	0.61	−0.25	1.00	
SiO ₄	−0.54	0.47	0.54	0.67	−0.22	−0.38	−0.42	0.19	−0.52	1.00

Bold correlations are significant at $p < 0.05$ & $N = 24$.

DO varied between a maximum of 8.2 mg/L during autumn at St.III and a minimum of 6.2 mg/L in summer at St.V. As a general trend, the lowest concentrations of DO that were detected during summer may be due to its consumption in the decomposition of detritus plankton and the complex organic matters. This result agrees with the fact that oxygen solubility decreases with increasing temperature and salinity as reported by Calliari et al. (2005).

Dissolved phosphate

The role of dissolved inorganic phosphorus could be considered as an important nutrient for marine phytoplankton in the oligotrophic settings and the need for evaluating nutrient limitation at the taxa and/or single cell level, rather than inferring it with nutrient concentrations and ratios or bulk enzyme activities (Mackey et al., 2007). The maximum concentration of dissolved phosphate (0.51 μM) was found during autumn at St.III which may be due to the effect of low sewage effluents and the land filing, while the minimum value (0.04 μM) was recorded during summer at St.V (Table 1), which is relatively far from the pollution sources.

Nitrate

Nitrate is the most stable form of inorganic nitrogen in the oxygenated waters. In the present study, dissolved nitrate attained a maximum of 2.27 μM during autumn at St.III, which sustained the highest abundance of the phytoplankton (average of 7071 unit/L) and a minimum of 0.07 μM during summer at the Mangrove station (St.V), coinciding with the lowest phytoplankton counts (average of 2770 unit/L). These observations are in accordance with those of El-Naggar et al. (2002), Nassar and Hamed (2003), Nassar (2007a) and Madkour et al. (2010) and confirmed by the significant positive correlation ($r = 0.86$) between the dissolved nitrate and phytoplankton counts in the study area.

Nitrite

Nitrite is one of the dissolved inorganic nitrogen forms present in water bodies and could be used as pollution indicator. It is not a stable end product, its absence or presence in such quantities might not be so peculiar (Collos, 1998). This could be obtained from the transformation to either nitrate or to ammonia. The nitrite accumulation in the water column is due to excretion by algal cells, which was estimated by

10–15% of the total amount of nitrogen entering the mixed-water column (Al-Qutob et al., 2002). The maximum nitrite concentration in the area of study (0.39 μM) was recorded during autumn at St.VI and the minimum (0.02 μM) was observed during summer at St.V.

Ammonium

Ammonium is the nitrogenous end product of the bacterial decomposition of natural organic matter containing nitrogen. In the presence of high ammonium concentrations, the phytoplankton productivity could be high or even higher if the cells are using NH_4^+ rather than NO_3^- (Dugadale et al., 2007). As shown in Table 1, the maximum concentration of ammonium (8.80 μM) occurred during summer at St.I, may be attributed to the effect of low sewage effluents of Al-Gemsha and the General Petroleum Companies, while the minimum (1.82 μM) was found during autumn at St.V, which is relatively far from pollution sources.

Reactive silicate

The reactive silicate displayed the highest concentration of 5.22 μM during summer at St.V, and the lowest of 0.41 μM during autumn at St.III. The low autumn silicate was associated with the high flourishing of dominant diatoms especially at St.III, explaining the inverse significant correlation between silicate and phytoplankton counts ($r = -0.54$) (Table 5). However, the irregular pattern of reactive silicate in the coastal waters could be attributed to the uptake of silica in the formation of diatom frustules (Wu and Chou, 2003). This agrees with the findings of Nassar and Hamed (2003), Toulbah et al. (2010) and Madkour et al. (2010).

Phytoplankton

Community composition and distribution

The phytoplankton community in the north western Red Sea was represented by 145 species including 81 Bacillariophyceae species, 45 of Dinophyceae, 11 Cyanophyceae, 6 Chlorophyceae and one species only for each of Euglenophyceae and Silicoflagellates. Diatoms were the most dominant group, since it constitutes about 76.4% of the total phytoplankton abundance with an average of 3654 cell/L, mainly due to the flourishing of *Climacosphenia moniligera* Ehrenberg (average of 213 cell/L), *Cylindrotheca closterium* (Ehrenberg) Reimann & J.C. Lewin

(average of 194 cell/L), *Chaetoceros simplex* Ostensfeld (average of 187 cell/L), *Chaetoceros curvisetus* Cleve (average of 165 cell/L), *Chaetoceros lorenzianus* Grunow and *Skeletonema costatum* (Greville) Cleve (average of 159 cell/L) as well as *Nitzschia pungens* var. *atlantica* Cleve (average of 119 cell/L) and *Licmophora flabellata* Greville (average of 101 cell/L) as shown in Table 4.

Dinophyceae was the second dominant group forming about 14.63% of the total phytoplankton with an average of 700 cell/L and a relative high occurrence of *Ceratium furca* (Ehrenberg) Claparède & Lachmann (average of 100 cell/L). Whereas, the other algal classes that include Cyanophyceae (4.12%), Chlorophyceae (3.95%), Euglenophyceae (0.21%) and Silicoflagellates (0.69%) were found in low counts, forming collectively about 9.0% of the total phytoplankton (Table 2 & Fig. 2).

Generally, recoding of 145 phytoplankton species in the present study is considered to be high as compared with the data reported by several workers on the Red Sea and some of the studied surrounding habitats, like the northern part of Suez Gulf known as Suez Bay (80 spp.; Nassar and Hamed, 2003), the northern part of the Red Sea (110 spp.; Shams El-Din et al., 2005), the sector of Halayib-Shalatin (25 spp.; Abel Rahman and Nassar, 2005), the Bitter Lakes and Tamsah Lake of the Suez Canal (116 spp.; Nassar and Shams El-Din, 2006), the western coast of the Gulf of Aqaba (127 spp.; Nassar, 2007b), Jeddah coast of Saudi Arabia Red Sea (73 spp.; Toulbah et al., 2010) and the Egyptian waters of eastern Mediterranean (88 spp.; El-Sherif et al., 2010). Whereas, about 181 phytoplankton species were observed in the Egyptian coast of the Red Sea (Madkour et al., 2010). However, a checklist of 207 phytoplankton species is detected in the Egyptian waters of the Red Sea and some surrounding habitats during the period 1990–2010 and represented by Nassar and Khairy (2014).

The highest abundance of phytoplankton was found at Hurgada-NIOF (St.III) with an average of 7070 unit/L followed by Al-Qusir (St.VI) with an average of 5283 unit/L and Al-Gemsha (St.I) with an average of 5177 unit/L. This may be due to the relatively high eutrophication state at these sites by subjecting to low sewage and oil effluents. However, the sewage discharge may increase the phytoplankton productivity where sewage was the main source of nitrogen and phosphorus (Burford et al., 2012). On the other hand, the lowest

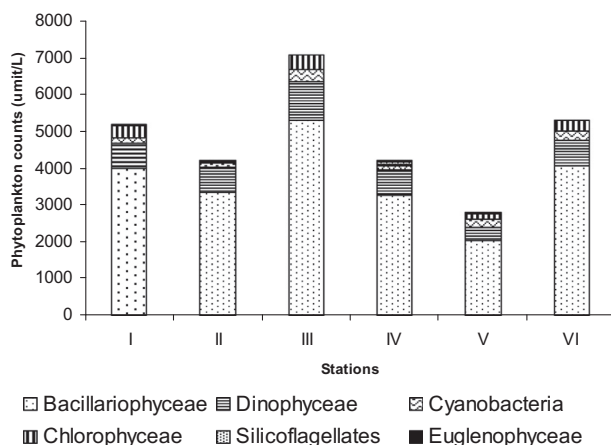


Figure 2 Annual average counts of the phytoplankton classes at the different sampling stations during 2013.

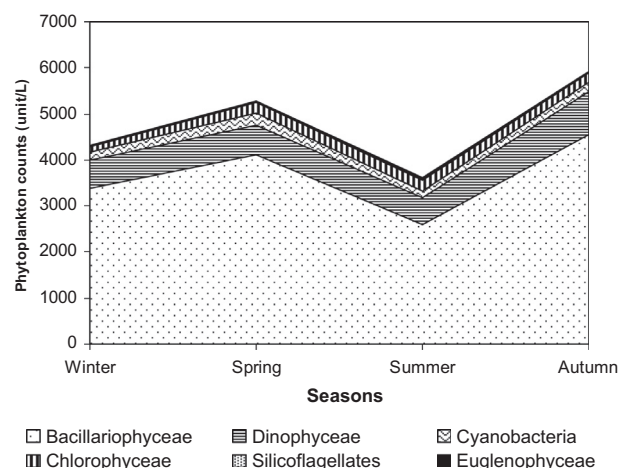


Figure 3 Seasonal variations of the different phytoplankton classes during 2013.

population density of phytoplankton was recorded at the mangrove, St.V (2770 unit/L) in accordance with Abel Rahman and Nassar (2005), who reported that the phytoplankton associated with mangroves was generally observed to have low counts of individuals and also low number of species. This phenomenon was also explained by Rajkumar et al. (2009) who included that the distribution and abundance of phytoplankton in tropical waters varied remarkably due to the environmental fluctuations during the seasons and these variations are well pronounced in the sheltered costal systems like mangroves.

Seasonal distribution

The total phytoplankton in the whole investigated areas showed the highest counts during autumn with an average of 5916 unit/L, followed by spring (average of 5282 unit/L) and winter (average of 4329 unit/L), while summer sustained the lowest counts (average of 3607 unit/L) as shown in Table 3 and Fig. 3. Generally, autumn was reported as the most productive season in the Red Sea by Levnon-Spanier et al. (1979), Khalil et al. (1984), Khalil and Ibrahim (1988), Nassar (2000) and Nassar and Hamed (2003). In the present study, the high abundance of phytoplankton during autumn was associated with high flourishing of the diatoms; *C. curvisetus* (average of 333 cell/L), *C. lorenzianus* (average of 273 cell/L) and *C. closterium* (average of 217 cell/L) and the dinoflagellate *C. furca* (Table 4). On the other hand, some phytoplankton species showed their maximum abundance during summer, which sustained the lowest abundance of phytoplankton. These taxa namely, *C. moniligera* (average of 460 cell/L) and *S. costatum* (average of 240 cell/L) of the diatoms and the chlorophyte *Chlorella salina* Butcher (average of 133 cell/L).

Statistical analysis

Correlation matrices and multiple regressions

The correlation matrices indicated that the total counts of phytoplankton was positively correlated with nitrate concentrations ($r = 0.86$), dissolved phosphate ($r = 0.60$) and the dissolved oxygen ($r = 0.55$), while it was inversely correlated

with water salinity ($r = -0.59$) and the reactive silicate ($r = -0.54$) as shown in Table 5.

The stepwise multiple regressions showed the high dependence of phytoplankton counts on the concentrations of nitrate and ammonium as well as reactive silicate and water salinity. These parameters were the most effective variables that controlled the seasonal fluctuations and species diversity of phytoplankton in the study area. The regression model was: $\text{phytoplankton counts} = 28564 + 0.69 \text{ NO}_3 + 0.284 \text{ NH}_4 - 0.13 \text{ SiO}_4 - 0.30 \text{ Salinity}$ (M.R. = 0.91, $N = 24$ and $p < 0.07$). This equation could be applied in the future to predict the total phytoplankton counts in the coastal waters of the northern part of the Red Sea, Egypt.

Similarity coefficient

The cluster analysis based on the seasonal fluctuations of phytoplankton counts at each site illustrated two groups at similarity level of $\sim 77\%$; the first group included stations V, III and VI, while the second one was further subdivided into two clusters at the similarity level of $\sim 83\%$; one of them represents St.I and the other includes stations II and IV (Fig. 4). The similarity dendrogram confirmed that the species composition and abundance of phytoplankton at the stations located South of Hurgada City differ greatly than that located in the North.

Species diversity

The diversity of phytoplankton organisms is important for the ecology and biogeochemistry of the ocean, and almost certainly lends stability to the system (Ptacnik et al., 2008). As shown in Fig. 5, the phytoplankton diversity index in the study area sustained high seasonal values, with averages of 3.81, 3.88, 3.36 and 3.97 during winter, spring, summer and autumn respectively, and varying between a maximum of 4.07 at St.III and a minimum of 3.49 at St.I. The high value at St.III may be attributed to the dominance of several phytoplankton species, while low diversity at St.I. was due to the clear dominance of the diatom; *C. moniliger* which formed about 43.5% of the total phytoplankton in summer at this station. These values are pronouncedly higher than those found in the Suez Bay

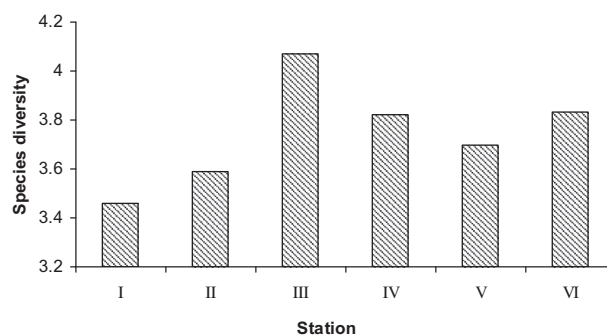


Figure 5 The species diversity at the different sites.

(average: 2.8, Nassar and Hamed, 2003) and Aqaba Gulf (average: 2.94, Nassar, 2007b).

References

- Abel Rahman, N.S., Nassar, M.Z., 2005. Preliminary studies on plankton communities associated with the mangrove forest habitats at Halayib-Shalatin Sector, Red Sea. Egypt. Pak. J. Mar. Sci. 14 (2), 133–144.
- Abou-Aisha, K.M., Kobbia, I.A., EL Abyad, M.S., Shabana, E.F., Schanz, F., 1995. Impact of phosphorus loadings on macro-algal communities in the Red Sea coast of Egypt. Wat. Air Soil Pollut. 83, 285–297.
- Al-Kandari M., Al-Yamani F., Al-Rifaie K., 2009. Marine Phytoplankton Atlas of Kuwait's Waters. Kuwait Institute for Scientific Research. ISBN 99906-41-24-2. pp. 351.
- AL-Najjar, T., Badran, M.I., Richter, C., Meyerthofer, M., Sommer, U., 2007. Seasonal dynamics of phytoplankton in the Gulf of Aqaba, Red Sea. Hydrobiologia 279, 69–83.
- AL-Qutob, M., Häse, C., Tilzer, M.M., Lazar, B., 2002. Phytoplankton drives nitrite dynamics in the Gulf of Aqaba, Red Sea. Mar. Ecol. Prog. Ser. 239, 233–239.
- American Public Health Association (APHA), 2005. Standard Methods for the Examination of Water & Wastewater, twenty first Ed. Washington.
- Armbrust, E.V., 2004. The genome of the diatom *Thalassiosira pseudonana*: ecology, evolution, and metabolism. Science 306, 79–86.
- Belkin, I.M., 2009. Rapid warming of large marine ecosystems. Prog. Oceanogr. 81, 207–213. <http://dx.doi.org/10.1016/j.pocean.2009.04.011>.
- Baars, M.A., Schalk, P.H., Veldhuis, M.J.W., 1998. Seasonal fluctuations in plankton biomass and productivity in the ecosystems of the Somali Current Gulf of Aden and Southern Red Sea. In: Sherman et al. (Eds.), Large Marine Ecosystems of the Indian Ocean: Assessment, Sustainability, and Management. Blackwell Science, Oxford, pp. 143–174. ISBN: 0632043180.
- Behrenfeld, M., O'Malley, R., Siegel, D., McClain, C., Sarmiento, J., Feldman, G., Milligan, A., Falkowski, P., Letelier, R., Boss, E., 2006. Climate-driven trends in contemporary ocean productivity. Nature 444, 752–755.
- Berge, T., Daugbjerg, N., Andersen, B.B., Hansen, P.J., 2010. Effect of lowered PH on marine phytoplankton growth rates. MEPS 416, 79–91.
- Bourrelly, P., 1968. Les algues d, eau douce, initiation a la systematique. In: Boubée, N., (Ed.), Tom II: Les algues Jaunes et Brunes, Chrysophycées, Phaeophycées, et Xanthophycées. Paris, pp. 438.
- Burford, M.A., Revill, A.T., Smith, J., Clementson, L., 2012. Effect of sewage nutrients on algal production, biomass and pigments in tropical tidal Creeks. Mar. Pollut. Bull. 64, 2671–2680.

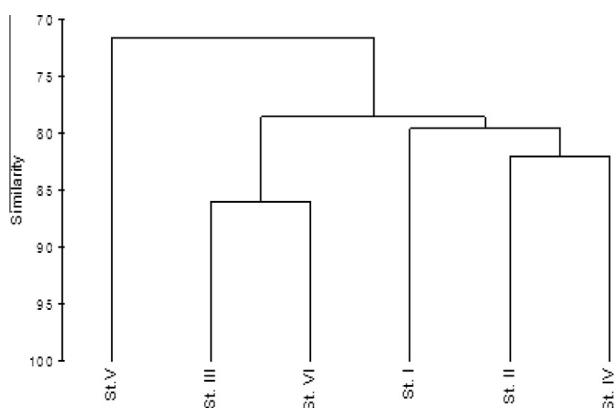


Figure 4 Bray Curtis of similarity of phytoplankton abundance between the different stations.

- Boyce, D., Lewis, M., Worm, B., 2010. Global phytoplankton decline over the past century. *Nat. News* 466, 591–596.
- Calliaria, D., Gomez, M., Gomez, N., 2005. Biomass and composition of the phytoplankton: large scale distribution and relationship with 166 environmental variables during a spring cruise. *Cont. Shelf Res.* 25, 197–210.
- Collos, Y., 1998. Nitrate uptake, nitrite release and uptake, and new production estimates. *Mar. Ecol. Prog. Ser.* 171, 293–301.
- Cupp, E.E., 1943. Marine plankton diatoms of the west coast of North America. University of California Press, Berkeley and Los Angeles, California, pp. 238.
- Deyab, M.A., Khedr, A.A., EL-Naggar, M.A., 2004. Phytoplankton distribution in relation to environmental factor along the Suez Canal and the Red Sea coast of Egypt. *Algol. Stud.* 112, 123–140.
- Dugadale, R.C., Wilkerson, F.P., Hogue, V.E., Marchi, A., 2007. The role of ammonium and nitrate in spring bloom development in San Francisco Bay. *EST Coast Shelf Sci.* 73, 17–29.
- El-Naggar, A.H., Osman, M.E.H., El-Sherif, Z.M., Nassar, M.Z., 2002. Phytoplankton and seaweeds of the western coast of Suez Gulf (from Red Sea) in relation to some physico-chemical factors, oil and sewage pollution. *Bull. Fac. Sci. Assiut* 31 (1), 77–104.
- EL-Nayal, A.A., 1935. Egyptian freshwater algae. *Bull. Fac. Sci. Cairo* (4), 106.
- El-Sherif, Z.M., Abo El-Ezz, S., 2000. Checklist of plankton of the northern Red Sea. *Pak. J. Mar. Sci.* 9 (1&2), 61–78.
- El-Sherif, Z.M., Nassar, M.Z., Fahmy, M.A., 2010. Phytoplankton distribution in the southeastern Mediterranean Sea (Egyptian waters) in summer and winter 2005. *Egypt. J. Aquat. Res.* 36 (4), 609–621.
- Ferguson Wood, E.J., 1968. Dinoflagellates of the Caribbean Sea and adjacent areas. Univ. of Miami press, Library of Congress Catalog card number 68–9512, 141.
- Field, J.G., Clarke, K.R., Warwick, R.M., 1982. A practical strategy for analyzing multispecies distribution patterns. *Mar. Ecol. Prog. Ser.* 8, 37–52.
- Ghazzawi, F.M., 1939. A study of Suez Canal plankters. The Phytoplankton Hydrobiology and Fish. Direct notes and memories, 24, pp. 83.
- Halim, Y., 1969. Plankton of the Red Sea. *Oceanogr. Mar. Biol. Annu. Rev.* 7, 231–275.
- Huber-Pestaluzzi, G., 1938. Das phytoplankton des Succwassers. I. Teile, Die Binnergewasser. Stuttgart, pp. 342.
- Khalil, A.N., Ibrahim, A.M.M., 1988. A seasonal study of the surface phytoplankton of the Red Sea, North of Jeddah. *Arab. Gulf. J. Sci. Res.* 6 (2), 189–204.
- Khalil, A.N., Khafagi, A.K., Ibrahim, A.M., 1984. Preliminary survey of micro- and macrophytes South of Jeddah, Red Sea. *Proc. Symp. Coral Reef Environ. Red Sea, Jeddah*, 555–576.
- Khomayis, H.S., 2002. The annual cycle of nutrient salts and chlorophyll-a in the coastal waters of Jeddah, Red Sea. *J. KAU: Mar. Sci.* 13, 131–145.
- Levnon-Spanier, I., Padan, E., Röss, Z., 1979. Primary production in a desert-enclosed Sea – Gulf of Eilat (Aqaba) Red Sea. *Deep Sea Res.* 26, 673–685.
- Longhurst, A.R., 2007. *Ecological Geography of the Sea*, Second Ed. Elsevier Inc., pp. 1–542, ISBN: 978-0-12-455521-1.
- Mackey, M.R.K., Labiosa, G.R., Street, H., Post, J., Paytan, A., 2007. Phosphorus availability, phytoplankton community dynamics, and taxon-specific phosphorus status in the Gulf of Aqaba, Red Sea. *Limnol. Oceanogr.* 52, 873–885.
- Madkour, F.F., El-Sherbiny, M.M., Amer, M.A., 2010. Phytoplankton population along certain Egyptian coastal regions of the Red Sea. *Egypt. Aquat. Biol. Fish.* 14 (2), 95–109.
- Mizuno, T., 1990. Illustrations of the Freshwater Plankton of Japan, ninth printing. Hoikush Publishing Co. Ltd., Japan, pp. 353.
- Nassar, M.Z., 2000. Ecophysiological studies on phytoplankton along the western coast of Suez Gulf. Philosophy Doctor Thesis, Faculty of Science, Tanta University.
- Nassar, M.Z., Shams El-Din, N.G., 2006. Seasonal dynamics of phytoplankton community in Bitter Lakes and Tamsah Lake. *Egypt. J. Aquat. Res.* 32 (1), 198–219.
- Nassar, M.Z., 2007a. Species composition and distribution of phytoplankton in the western coast of Suez Gulf. *Egypt. J. Aquat. Res.* 33 (2), 113–132.
- Nassar, M.Z., 2007b. Nutrients and phytoplankton distribution in the coastal waters of Aqaba Gulf, Red Sea. *Egypt. J. Aquat. Res.* 33 (2), 133–151.
- Nassar, M.Z., Hamed, M.A., 2003. Phytoplankton standing crop and species diversity in relation to some water characteristic of Suez Bay (Red Sea). *Egypt. J. Aquat. Biol. Fish.* 7 (3), 25–48.
- Nassar, M.Z., Khairy, H.M., 2014. Checklist of phytoplankton species in the Egyptian waters of the Red Sea and some surrounding habitats (1990–2010). *J. Ann. Res. Rev. Biol.* 4 (23), 3566–3585.
- O'Connor, M.I., Piehler, M.F., Leech, D.M., Anton, A., Bruno, J.F., 2009. Warming and resource availability shift food web structure and metabolism. *PLoS Biol.* 7 (8).
- Paeerl, H.W.L., Valdes, L.M., Pinckney, J.L., Piehler, M.F., Dyble, J., et al., 2003. Phytoplankton photopigments as indicators of estuarine and coastal eutrophication. *Bioscience* 53, 953–964.
- Peragallo, H., Peragallo, M., 1908. Diatomees marines de France et des Districts Maritimes Voisons, I–III (text and plates). Paris and Grez Sur-Loing, pp. 1–491.
- Prescott, G.W., 1962. Algae of the western Great Lakes area. Brown, W. C., Dubuque (IOWA), pp. 977.
- Ptácnik, R., Solimini, A.G., Andersen, T., Tamminen, T., Brettum, P., Lepistö, L., Willén, Rekolainen, S., 2008. Diversity predicts stability and resource use efficiency in natural phytoplankton communities. *Proc. Nat. Acad. Sci.* 105, 5134–5138.
- Qurban, M.A., Balala, A.C., Kumar, S., Bhavya, P.S., Wafar, M., 2014. Primary production in the northern Red Sea. *J. Mar. syst.* 132, 75–82.
- Rajkumar, M., Perumal, P., Prabu, A.V., Perumal, N.V., Rajeskar, K.T., 2009. Phytoplankton diversity in Pichavaram mangrove waters from South-east coast of India. *J. Environ. Biol.* 30, 489–498.
- Schumann, R., Baudler, H., Glass, A., Dümcke, K., Karsten, U., 2006. Long-term observations on salinity dynamics in a tide less shallow coastal lagoon of the southern Baltic Sea coast and their biological relevance. *J. Mar. Syst.* 60, 193–404.
- Shakweer, L., 2003. Ecological and fishery investigations of Nozha Hydrome near Alexandria 2000–2001. 1. Chemistry of Nozha Hydrome water under the conditions of fertilizers applications. *Bull. Natl. Inst. Oceanogr. Fish.* 29, 387–425.
- Shannon, C.E., Wiener, 1963. The mathematical theory of communications. Univ. Illinois, Urbana, pp. 117.
- Shams-El-Din, N.G., Nassar, M.Z., Abd-El-Rahmann, N.S., 2005. Surveillance studies on plankton in the northern part of the Red Sea during winter and summer, 2002. *J. Egypt. Ger. Soc. Zool.* 48 (D).
- Shaikh, E.A., Roff, J.C., Dowidar, N.M., 1986. Phytoplankton ecology and production in the Red Sea off Jeddah, Saudi Arabia. *Mar. Biol.* 92, 405–416.
- Sofianos, S.S., Johns, W.E., 2002. An oceanic general circulation model (OGCM) investigation of the Red Sea circulation, 1. Exchange between the Red Sea and the Indian Ocean. *J. Geophys. Res.* 107, 3196. <http://dx.doi.org/10.1029/2001JC001184>.
- Sommer, U., 2000. Scarcity of medium-sized phytoplankton in the northern Red Sea explained by strong bottom – up and weak top – down control. *Mar. Ecol. Prog. Ser.* 197, 19–25.
- Sourina, A., 1986. Atlas Du Phytoplankton Marin, Introduction, Cyanophycees, Dictyochophycees, Dinophycees et Radiophycees, vol. 1, pp. 21.
- Sridhar, R., Thangaradjou, T., Kumar, S.S., Kannan, L., 2006. Water quality and phytoplankton characteristics in the Palk bay, south-east coast of India. *J. Environ. Biol.* 27, 561–566.
- Stewart, K.D., Mattox, K.R., 1975. Comparative cytology evolution and classification of green algae with some consideration of the other organisms with chlorophylls a and b. *Bot. Rev.* 41, 104–135.

- Strickland, J.D.H., Parsons, T.R., 1972. A manual of sea water analysis. Can. Fish. Res. Board Bull. 167, 310.
- Toulibah, H.E., Abu EL-Kheir, W.S., Kuchari, M.G., Abdulwassi, N.H., 2010. Phytoplankton composition at Jeddah Coast - Red Sea Saudi Arabia in relation to some ecological factors. JKAU: Sci. 22, 115–131.
- Wu, J.T., Chou, T.L., 2003. Silicate as the limiting nutrient for phytoplankton in a subtropical eutrophic estuary of Taiwan. Estuar. Coast. Shelf Sci. 58, 155–162.
- Utermöhl, H., 1936. Quantitative methoden zur untersuchung des nannoplankton. Adderheldens Handbuch der Biolog. Arb. Methoden IX (2), 1879–1937.